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(71) Applicant *(for all designated States except US)*: FOXWOOD RESEARCH LIMITED [GB/GB]; c/o Nicolas and Walters, 54/56 Victoria Street, Shirebrook, Mansfield, Nottinghamshire NG20 8AQ (GB).

(72) Inventors; and
(75) Inventors/Applicants *(for US only)*: BLOWES, Phillip, Charles [GB/GB]; Carema, Timberidge, Rickmansworth WD3 4JD (GB). TAYLOR, Alan, John [GB/GB]; Greenacres, Inkersall Green Road, Inkersall, Chesterfield, Derbyshire S43 3HA (GB). ROBERTS, George [GB/GB]; Nicolas and Walters, 54/56 Victoria Street, Shirebrook, Mansfield, Nottinghamshire NG20 8AQ (GB). WOOD, Fran [GB/GB]; Nicolas and Walters, 54/56 Victoria Street, Shirebrook, Mansfield, Nottinghamshire NG20 8AQ (GB).

(74) Agent: SAUNDERS & DOLLEYMORE; 9 Rickmansworth Road, Watford, Hertfordshire WD1 7HE (GB).

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(54) Title: SUBSTRATES WITH BIOCIDAL PROPERTIES AND PROCESS FOR MAKING THEM

(57) Abstract

The invention relates to a process for treating a substrate to impart biocidal properties to the substrate by depositing or impregnating the substrate with solubilised chitosan, immersing the substrate in a solution of a silver salt, treating the substrate to reduce the silver salt to atomic/metallic silver, cross–linking the chitosan and washing the resultant treated substrate. The invention also relates to biocidal substrates when prepared by this process and to the use of such biocidal substrates for the manufacture of garments, dressings and protective drapes suitable for medical and veterinary use.

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SUBSTRATES WITH BIOCIDAL PROPERTIES AND PROCESS FOR MAKING THEM

This invention relates to a treatment process suitable for application to fibre, yarn, non-woven, woven, knitted fabric, garments, or paper products, hereinafter referred to as 'The Substrate'. It also relates to articles fabricated from these materials, in particular those widely used in the health care and associated industries. The treatment process can be applied to any natural or synthetic substrate or blends therefrom. Typically, cotton, wool, viscose, polyamide, polyester, and/or polypropylene fibres or mixtures may be employed. Substrate type and specification are chosen in accordance with the end-use.

The invention imparts a characteristic that effectively kills a wide range of micro-organisms which come into contact with the treated material. It also prevents decomposition of the underlying substrate by inactivating or killing bacteria and other microbes on contact or in solution. The resultant treatment is partly or wholly resistant to degradation caused by most laundering and steam sterilisation processes. The treatment provides a characteristic coloration and some stiffening to the substrate, and improves the removal of organic staining when washed in a commercial laundry process.

It is known that atomic/metallic silver is a highly effective contact biocide, and chitosan is also known to exhibit the property of inactivating a wide range of micro-organisms which come into contact with it. It has now been found that atomic/metallic silver can be effectively dispersed through a polymeric structure of chitosan which itself has been deposited on the substrate. The dispersion is such that the silver substantially or wholly retains its biocidal properties. It has also been found that the system retains its biocidal properties when the chitosan is rendered insoluble and is fixed to the substrate. It has been shown that chemicals do not leach out from this system to any significant degree over the pH range of 5-9.

According to one aspect of the invention there is provided a process for treating a natural or synthetic substrate or blends thereof to impart biocidal properties to the substrate, said process comprising the steps of: (a) depositing or impregnating the substrate with solubilised chitosan, (b) immersing the substrate in a solution of silver salt, (c) treating the substrate to reduce the silver salt to atomic/metallic silver, (d) cross-linking the chitosan, and (e) washing and drying the resultant treated substrate.

According to a second aspect of the invention there is provided a biocidal substrate comprising a natural or synthetic, woven, non-woven or knitted fabric impregnated with atomic/metallic silver bound to cross-linked chitosan polymer.

The atomic/metallic silver is bound into the chitosan polymer by interaction with the amine groups of the latter, and is further entrapped by the cross-linking process which renders the chitosan insoluble. Silver is not easily removed from the system and represents a significant improvement on previous systems that merely utilise surface adsorption to provide a carrier for the silver. Such previous systems may also result in a "dusty" substrate surface, and a substrate that is difficult to handle and launder. The process of the present invention may be applied to substrates with a wide variety of end-uses, changing the substrate properties only marginally except with regard to anti-microbial properties.

The chitosan, which is required to have a minimum level of deacetylation sufficient to impart acid solubility, may be applied by deposition on, or impregnation of the substrate and rendered insoluble over a wide range of pH in conditions applicable to the use of surgical and similar garments, face masks, and health care dressings. The attachment of the chitosan to the substrate fibres is strong so that the resulting chitosan treatment can be regarded as very durable, often lasting the lifetime of the fabricated article and able to withstand

commercial processes such as laundering and sterilisation.

Oxidative bleaching if required must be carried out on the substrate prior to treatment with chitosan. The treatment process imparts colour to the finished product which is characteristic of the process. Staining of articles in use is reduced due to the chemical nature of chitosan, which binds organic chromophores under acidic pH and releases them in alkaline conditions, as is used in commercial laundering processes. Stains are therefore washed from the treated fabric with greater ease than the untreated substrate.

The extent of treatment in respect to quantity of silver and chitosan to be deposited on the substrate is determined by the degree of biocidal effect required in the end product. It is also varied according to the nature of the substrate. Typically a deposition of 0.5 – 3% of chitosan and 0.01 – 2% of silver salt by weight of fabric is suitable for end-use as garments and face masks.

Chitosan is solubilised by any of the methods common in the art, typically using a 2% w/w solution of acetic acid and may be applied to the substrate by padding, printing, spraying or pressure impregnation, and squeezed in a commercial mangle. The pressure of squeeze is adjusted to provide the amount of chitosan deposition required for ultimate performance of the end-use article. The chitosan is applied using one or several passes through the padder/mangle.

After neutralising the treated substrate with aqueous alkali such as caustic soda, it is immersed in a solution of silver salt, typically the nitrate, of strength to suit the ultimate end-use of the system, generally in the range 0.0005 – 2.0% w/v, preferably 0.001 – 0.2% w/v, and is then squeezed in a commercial mangle to remove excess liquid. Whilst silver nitrate is the preferred salt, any water soluble silver salt such as the acetate, perchlorate, difluoride, lactate or propionate may be used.

The neutralised substrate is then treated to reduce the silver salt to atomic/ metallic silver by one of the methods common in the art, of which visible and/or ultraviolet light, gaseous hydrogen or hydroquinone are preferred for reasons of practicality.

The neutralised substrate can be bathed in an aqueous mixture of 0.2% w/v hydroquinone and 0.5% w/v sodium carbonate for up to 4 hours. Alternatively, one or more passes under a bank of commercial visible and/or ultraviolet light sources can be used to effect full conversion to atomic/metallic silver. The photochemical reduction process is made more efficient by a preliminary treatment in a 1% w/v solution of sodium chloride or a sodium chloride/sodium bromide mixture. If gaseous hydrogen is used the silver salt is first converted to the oxide using aqueous alkali before being reduced by hydrogen.

After washing with clean water the chitosan is cross-linked to render it insoluble at acidic pH. The cross-linking is performed using any of the methods common in the art, for instance by contacting the substrate with glutaraldehyde at a strength of 0.01 – 2.0% w/v for a period of 1-24 hours at room temperature. Alternatively, other dialdehydes or epichlorhydrin may be used.

The substrate is then washed with clean water and dried and is then ready to be fabricated into articles for end-use in medical and veterinary medicine such as face masks, dressings, surgical drapes, surgical gowns, protective clothing, as well as incontinence pads, sanitary towels, nappies, gloves, protective wrappings, sterile field and filters suitable for air, water or blood filtration.

Alternatively, the cross-linking process may be carried out before immersion in the silver salt solution. Here the chitosan-treated substrate is neutralised with aqueous alkali and the cross-linking process applied as described above.

After washing with clean water, the substrate is immersed in silver salt solution, squeezed and reduced as previously described. After washing with clean water, the substrate is dried and is ready for fabrication into end-uses.

Alternatively the cross-linking process may be modified to render the chitosan partially soluble at acidic pH. Here a low concentration of the cross-linking chemical is used, for instance glutaraldehyde, at a strength of 0.001 – 0.01% w/v, the amount being determined by reference to the quantity of chitosan deposited on the substrate and designed to cross-link a known proportion of the chitosan, for instance 50%. This enables a slow release of the silver and chitosan into the acidic aqueous phase while maintaining the contact biocide properties of the system itself.

In some instances it is possible to combine the steps of chitosan cross-linking with the silver salt treatment.

Generally, biocidal activity is diminished in use, not by significant use of the active ingredients (silver and chitosan), but by the accumulation of biomass which may be bound to the material, e.g. by a process similar to chelation. The material may be reactivated by washing at alkaline pH with clean water, and preferably with a cationic surfactant present. If the material has been in use under slightly or moderately alkaline conditions, washing at acidic pH and preferably with an anionic surfactant will effect reactivation wholly or in part.

The invention may be illustrated with reference to the following examples:

EXAMPLE 1

Method

Two samples of scoured 300g/m^2 cloth, weft 60 picks/inch (cotton) warp;

94 ends/inch (polyester) typically used for work-wear apparel, were treated with chitosan and silver and subjected to a microbial challenge test followed by a quantitative estimation of survivors in accordance with standard SNV 195 924 (Textile Fabrics: determination of the antibacterial activity, germ count method). (SNV 195 924 is a Swiss testing system protocol for textiles from Schweizerischen Normen-Vereinigung, Kirchen Veg 4, Postfach 8032, Zurich.)

Treatment system

AT2/B – coated with 1.5%w/w of chitosan, neutralised and immersed in a 0.02%w/v solution of silver nitrate for 8 hours, washed with distilled water and placed in a bath of 1% sodium chloride for 1 hour. After washing, the cloth was cross-linked by immersing in 0.1w/v aqueous solution of glutyraldehyde for 8 hours and placed under a 20 watt 600 mm tube ultraviolet light source for 15 minutes and dried in air.

AT2/C – as sample AT2/B, with the sodium chloride step omitted.

Microbial challenge

Samples were flash sterilised at 1.67 barg and 115°C for 1 minute and cut into circles 4.5cm diameter, three pieces per sample, and placed in screw-top jars which had also been previously flash sterilised to the same conditions.

An overnight culture of staphylococcus aureus NCTC 10788 was diluted aseptically in tryptone soya broth to give an approximate concentration of 10^3 cells per ml.

1ml of this suspension was added to each of the samples, as far as possible covering the whole surface of the sample, the volume chosen to ensure no free liquor remained, i.e. all microbes were kept in contact with the samples.

The jars were incubated at 30°C for six hours whilst shaking at 100rpm on an orbital shaker.

Testing for anti-microbial activity:

The samples were transferred aseptically to container of 10 ml of tryptone soya broth and vortexed vigorously. This process is intended to remove any bacteria from the fabric. The samples were removed as quickly as practical, squeezed in a clean empty dish, and placed firmly onto the surface of a fresh tryptone soya agar plate. Growth here would indicate the presence of live bacteria clinging to the sample. Results are shown in the table below "Beneath washed sample".

The broths vortexed from the samples were serially diluted down to 10^{-4} dilution and plate counts made of each dilution by the surface spread method. These counts give a quantitative estimation of cell numbers of live bacteria washed from the samples. Results shown in the table under "Plate count-washings".

These diluted broths were incubated for 24 hours at 37°C and examined by eye to determine presence or absence of turbidity. Turbidity indicates the survival of just one or two bacteria which may have been missed in the plate count of the diluted broths. Findings are reported in the table as "Incubated washings".

Results

Results of microbiology are given in the following table, average values for three test pieces of each sample:

	Sample	AT2/B	AT2/C
Plate count-Washings	10°	0	0
	10^{-1}	0	0
	10^{-2}	0	0
	10^{-3}	0	0
	10^{-4}	0	0
Incubated Washings	10°	Growth	Growth
	10^{-1}	No growth	No growth
	10^{-2}	No growth	No growth
	10^{-3}	No growth	No growth
	10^{-4}	No growth	No growth
Beneath washed sample		No growth	No growth

Conclusion

The results strongly indicate that samples AT2/B and AT2/C are virtually or probably 100% effective as biocides against staphylococcus aureus.

EXAMPLE 2

Method

Two samples of cloth of the same specification as in Example (1) were treated in the following manner:

AT3/B6 – coated with 1% w/w of chitosan, neutralised and cross-linked using 0.02%w/v aqueous solution of glutyraldehyde, washed with clean water and immersed in a 0.005%w/v solution of silver nitrate for 8 hours. It was then steeped for 60 minutes in a mixture of 0.4%w/v aqueous solution of sodium chloride and 0.4% w/v aqueous solution of sodium bromide. After washing the treated cloth was placed under a 20 watt 600 mm tube ultraviolet light source for 15 minutes and dried in air.

AT3/F10 – immersed in a 0.25%w/v solution of silver nitrate for 8 hours. It was then steeped for 60 minutes in a mixture of 0.4w/v aqueous solution of sodium chloride and 0.4%w/v aqueous solution of sodium bromide. After washing the treated cloth was placed under a 20 watt 600 mm tube ultraviolet light source for 15 minutes and dried in air. (i.e. chitosan coating and cross-linking steps omitted).

Both samples were washed in an automatic industrial laundry cycle using a commercial washing powder and dried in an industrial drier. Each sample was washed once at 60°C and twice at 95°C and dried.

The samples were tested using the strain of the micro-organism staphylococcus aureus designated NCTC 10788. The test protocol was the same as example (1), but dilutions were performed to 10^{-3}. Results, averaged over three pieces for each sample, are given in the table below:

Sample Reference		AT3/B6	AT3/F10
Treatment system		*Chitosan, X-linked* *Silver*	*Silver*
Plate count- Washings	10° 10^{-1} 10^{-2} 10^{-3}	0 0 0 0	31 9 2 0
Incubated Washings	10° 10^{-1} 10^{-2} 10^{-3}	No growth No growth No growth No growth	Growth Growth No growth No growth
Beneath washed sample		No growth	A few colonies on edges, no growth underneath.

Conclusions

Sample AT3/B6 indicates extremely good anti-microbial activity and sample AT3/F10 shows probable significant anti-bacterial activity. After laundering three times in severe conditions, the chitosan treatment system shows a higher degree of biocidal activity than the silver-only treatment.

EXAMPLE 3

Method

Samples of cloth described in example (1) were treated in the following ways and then tested in accordance with the SNV 195 924 protocol described in example (1) using the micro-organism staphylococcus aureus NCTC 10788. Results of these tests are summarised in the table, all results being the average for three test pieces per sample.

AT3/20 – this sample was not treated and used as a control.

AT2/D – 1%w/w of chitosan was padded on to the cloth, neutralised and then cross-linked in a 0.02%w/v aqueous solution of glutyraldehyde for 8 hours. The cloth was washed, dried in air and tested.

AT3/F18 – the cloth was immersed in a 0.005%w/v solution of silver nitrate for 8 hours. It was then steeped for 60 minutes in a mixture of 0.4%w/v aqueous solution of sodium chloride and 0.4%w/v aqueous solution of sodium bromide. After washing the treated cloth was placed under a 20 watt 600 mm tube ultraviolet light source for 15 minutes and dried in air and tested.

AT2/A – 1%w/w of chitosan was padded on to the cloth, neutralised and then cross-linked in a 0.02%w/v aqueous solution of glutyraldehyde for 8 hours. After rinsing with clean water the sample was then treated as AT3/F18 sample.

Results

Sample Reference		AT3/20	AT2/D	AT3/F18	AT2/A
Treatment summary		*None*	*Chitosan* *Cross-linked*	*Silver*	*Chitosan* *Cross-linked* *Silver*
Plate count-Washings	10°	>300	>300	33	0
	10^{-1}	>300	>300	8	0
	10^{-2}	>300	>300	2	0
	10^{-3}	61	>300	0	0
	10^{-4}	32	256	0	0
Incubated Washings	10°	Growth	Growth	Growth	Growth
	10^{-1}	Growth	Growth	Growth	No growth
	10^{-2}	Growth	Growth	No growth	No growth
	10^{-3}	Growth	Growth	No growth	No growth
	10^{-4}	Growth	Growth	No growth	No growth
Beneath washed sample		Massive Growth	Growth	No growth	No growth

Conclusions

Sample reference AT3/20 displayed an absence of anti-bacterial action. Sample AT2/D showed signs of inhibition, AT3/F18 is probably significantly biocidal and AT2/A is probably 100% effective biocidal activity. The combination of cross-linked chitosan with silver is more effective than silver or chitosan on their own.

EXAMPLE 4

Method

Two samples of the cloth referenced in example 3 were treated and tested to SNV 195 924 (see example (1) for details) using two micro-organisms, staphylococcus aureus NCTC 10788 and salmonella typhimurium strain NCTC 74.

Treatment: the samples were coated with 1% w/w of chitosan, neutralised and cross-linked using 0.02%w/w aqueous solution of glutyraldehyde for 8 hours, washed with clean water and immersed in a 0.005%w/v solution of silver nitrate for 1 hour. It was steeped for 5 minutes in a 1.0%w/v solution of sodium chloride. The treated cloth washed and was placed under a 20 watt 600 mm tube ultraviolet light source for 15 minutes and dried in air and tested.

Post treatment: the samples were laundered in an industrial washing cycle, once at 60°C and twice at 95°C and dried in a tumble drier.

Results

Sample Reference		AT3/A9	AT3/A9
Micro-organism		*Staph. Aureus*	*Salmonella typh.*
Plate count-Washings	10°	2	35
	10^{-1}	0	6
	10^{-2}	0	0
	10^{-3}	0	0
Incubated Washings	10°	No growth	No growth
	10^{-1}	No growth	No growth
	10^{-2}	No growth	No growth
	10^{-3}	No growth	No growth
Beneath washed sample		No growth	No growth

Conclusion

The samples are extremely biocidal to the micro-organisms used in the test.

EXAMPLE 5

Samples of cotton cloth, 230 g/m^2 (weft 74 picks/inch, warp 56 ends/inch) were treated with either 0.8%w/w or 1.5%w/w chitosan and cross linked using 0.4%w/v glutyraldehyde for 12 hours, washed and neutralised. They were tested according to test method SNV 195 924 Textile Fabrics: determination of the antibacterial activity: germ count method.

Incubation temperatures were 37°C and samples were flash sterilised at the start of the test at 115°C and 1.67 barg pressure.

At the end of 6 and 12 hours, 100 ml of sterile distilled water containing 3% Tween 80 as neutraliser was added aseptically to the jars containing the samples, shaken vigorously for 1 minute and 1 ml aliquots removed for plating out.

Three micro-organisms were used in the tests:

methicillin resistant staphylococcus aureus strain SDRU R80/606
klebsiella pneumoniae strain NCIMB 10341
salmonella typhimurium strain NCTC 74

The $\log_{10}$ CFU (colony forming units) figures were calculated and are tabulated below:

Results

Sample	MRSA			Kleb. Pneumoniae			Salmonella typh.		
Control	5.81	9.05	>10.5	6.16	9.13	>10.5	6.33	>9.5	10.11
0.8%	4.58	7.01	8.36	4.88	6.85	8.70	5.19	8.32	8.91
1.5%	4.94	6.93	7.85	4.99	7.16	8.96	5.01	9.25	9.32

Conclusions

Both samples showed some growth inhibition of all the microbes tested, but do not pass the criteria for antibacterial action. The treatment system using chitosan alone is not effective as a contact biocide.

CLAIMS

1. A process for treating a natural or synthetic substrate or blends thereof to impart biocidal properties to the substrate, said process comprising the steps of:

(a) depositing or impregnating the substrate with solubilised chitosan,

(b) immersing the substrate in a solution of silver salt,

(c) treating the substrate to reduce the silver salt to atomic/metallic silver,

(d) cross-linking the chitosan, and

(e) washing the resultant treated substrate.

2. The process according to Claim 1, wherein the step of cross-linking the chitosan is carried out before the substrate is immersed in the silver salt solution.

3. The process according to Claim 1, wherein the steps of cross-linking the chitosan is carried out at the same time as the silver salt solution is applied to the substrate.

4. The process according to any one of Claims 1 to 3, wherein the substrate is comprised of fibres or yarn, is a non-woven, woven or knitted fabric or garment or a paper product.

5. The process according to Claim 4, wherein the fibres or yarn are cotton, wool, viscose, polyamide, polyester or polypropylene or mixtures thereof.

6. The process according to any one of Claims 1 to 4, wherein the treated substrate comprises from 0.5 to 3% by weight chitosan and from 0.01 to 2% by weight silver salt.

7. The process according to any one of Claims 1 to 6, wherein the chitosan is solubilised in dilute acid.

8. The process according to Claim 7, wherein after treating the substrate with chitosan and before immersing said substrate in the silver salt solution the chitosan-treated substrate is neutralised with aqueous alkali.

9. The process according to any one of Claim 1 to 8,wherein the solubilised chitosan is applied to the substrate by padding, printing, spraying or pressure impregnation.

10. The process according to any one of Claims 1 to 9, wherein the solution of silver salt is a solution of silver nitrate or any water soluble silver salt such as the acetate, perchlorate, difluoride, lactate, or propionate salt.

11. The process according to Claim 10, wherein the concentration of silver nitrate solution is from 0.0005 to 2.0% w/v.

12. The process according to Claim 11, wherein the concentration of silver nitrate solution is from 0.001 to 0.2% w/v.

13. The process according to any one of Claims 1 to 12, wherein the silver salt is converted to the oxide by aqueous alkali and reduced by gaseous hydrogen.

14. The process according to any one of Claims 1 to 12, wherein the silver salt is reduced by visible and/or ultraviolet light.

15. The process according to Claim 14, wherein the substrate is treated with a solution of dilute sodium chloride or a sodium chloride/sodium bromide mixture prior to reducing the silver salt by visible and/or ultraviolet light.

16. The process according to any one of Claims 1 to 12, wherein the silver salt is reduced by hydroquinone.

17. The process according to Claim 16, wherein the silver salt is reduced using an aqueous mixture of 0.2% w/v hydroquinone and 0.5% w/v sodium carbonate.

18. The process according to any one of Claims 1 to 17, wherein the chitosan is cross-linked by contacting the chitosan-treated substrate with an aqueous solution of glutaraldehyde, or other dialdehyde or epichlorhydrin.

19. The process according to Claim 18, wherein the concentration of glutaraldehyde is from 0.001 to 0.1% w/v.

20. The process according to Claim 18, wherein the concentration of glutaraldehyde is from 0.01 to 2.0% w/v.

21. The process according to any one of Claims 18 to 20, wherein the chitosan-treated substrate is contacted with glutaraldehyde for a period of 1 to 24 hours at room temperature.

22. A biocidal substrate when prepared according to the method as claimed in any one of Claims 1 to 21.

23. A biocidal substrate comprising a natural or synthetic, woven, non-woven or knitted fabric impregnated with atomic/metallic silver bound to cross-linked chitosan polymer.

24. Use of the biocidal substrate according to Claim 22 or Claim 23 for the manufacture of face masks, dressings, incontinence pads, drapes, surgical gowns, protective clothing, sanitary towels, nappies, gloves, protective wrappings, sterile field or filters.

INTERNATIONAL SEARCH REPORT

Inte .ional Application No
PCT/GB 00/00604

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 D06M16/00 D06M15/17 D06M15/03 D06M11/83 A61L15/46
A01N59/16 D21H21/36

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 D06M A61L A01N D21H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE WPI Section Ch, Week 199549 Derwent Publications Ltd., London, GB; Class D22, AN 1995-378809 XP002138834 & JP 07 256025 A (NAKAMURA K), 9 October 1995 (1995-10-09) abstract ---	23,24
A	EP 0 291 587 A (SHIRLEY INST) 23 November 1988 (1988-11-23) page 3, column 3, line 23 - line 30; claims ---	1,4,10, 18,21-24
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[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

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Date of the actual completion of the international search	Date of mailing of the international search report
26 May 2000	09/06/2000

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL – 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Blas, V

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INTERNATIONAL SEARCH REPORT

Inte. .onal Application No
PCT/GB 00/00604

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication,where appropriate, of the relevant passages	Relevant to claim No.
A	DATABASE WPI Section Ch, Week 199651 Derwent Publications Ltd., London, GB; Class A60, AN 1996-514846 XP002138835 & JP 08 268821 A (SANGI KK), 15 October 1996 (1996-10-15) abstract	1,4,5,7, 10,22-24
A	DATABASE WPI Section Ch, Week 199641 Derwent Publications Ltd., London, GB; Class D22, AN 1996-406189 XP002138836 & JP 08 196461 A (NAKAMURA K), 6 August 1996 (1996-08-06) abstract	1,4,5,9, 22-24
A	DATABASE WPI Section Ch, Week 199629 Derwent Publications Ltd., London, GB; Class C07, AN 1996-283363 XP002138837 & JP 08 119805 A (NAKAMURA K), 14 May 1996 (1996-05-14) abstract	1,4,5,9, 22-24
A	DATABASE WPI Section Ch, Week 198045 Derwent Publications Ltd., London, GB; Class A60, AN 1980-79570C XP002138838 & JP 55 122556 A (NIPPON TENNEN GAS K), 20 September 1980 (1980-09-20) abstract	1,4,5,7, 22-24
A	US 5 643 971 A (ROENIGK KARL F) 1 July 1997 (1997-07-01) column 2, line 64 -column 3, line 21; claims	1,22-24

INTERNATIONAL SEARCH REPORT

Information on patent family members

Inte. .ional Application No
PCT/GB 00/00604

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
JP 7256025 A	09-10-1995	NONE	
EP 0291587 A	23-11-1988	EP 0460774 A	11-12-1991
		GB 2182560 A,B	20-05-1987
		US 4960413 A	02-10-1990
JP 8268821 A	15-10-1996	NONE	
JP 8196461 A	06-08-1996	NONE	
JP 8119805 A	14-05-1996	NONE	
JP 55122556 A	20-09-1980	JP 1148797 C	26-05-1983
		JP 57041950 B	06-09-1982
US 5643971 A	01-07-1997	US 5541233 A	30-07-1996
		US 5821271 A	13-10-1998
		AU 4921997 A	30-04-1998
		AU 682832 B	23-10-1997
		AU 5453294 A	22-06-1994
		BR 9307554 A	25-05-1999
		CN 1091140 A	24-08-1994
		DE 69319921 D	27-08-1998
		DE 69319921 T	15-04-1999
		EP 0671882 A	20-09-1995
		JP 8503941 T	30-04-1996
		MX 9307319 A	29-07-1994
		NO 952163 A	01-06-1995
		NZ 257762 A	24-03-1997
		WO 9412034 A	09-06-1994